eluent. 3α -O-Butanoyldeoxycholic acid, methyl ester was obtained in 85% yield. The product was crystallized from diethyl etherhexane: mp 110-2 °C. ¹H NMR: δ 0.60 (3 H, s, C-18 Me), 0.86 $(3 \text{ H}, \text{ s}, \text{C-19 Me}), 3.94 (1 \text{ H}, \text{ t}, J = 2.8 \text{ Hz}, \text{H-12}\beta), 4.67 (1 \text{ H}, \text{sept})$ $J_1 = 10.6$ Hz, $J_2 = 4.9$ Hz, H-3 β). Anal. Calcd for $C_{29}H_{48}O_5$: C, 73.47; H, 10.20. Found: C, 73.75; H, 10.03.

The acylations of 12, 13, and 14 were carried out similarly. The products were recovered (yields between 70 and 78%) and characterized.

 3α -O-Butanoylchenodeoxycholic acid, methyl ester: oil. ¹H NMR; δ 0.62 (3 H, s, C-18 Me), 0.88 (3 H, s, C-19 Me), 3.83 (1 H, q, J = 2.5 Hz, H-7 β), 4.56 (1 H, sept, $J_1 = 10.6$ Hz, $J_2 =$ 5 Hz, H-3β). Anal. Calcd for C₂₉H₄₈O₅: C, 73.47; H, 10.20. Found: C, 72.71; H, 9.80.

 3α -O-Butanoylursodeoxycholic acid, methyl ester: oil. ¹H NMR: δ 0.65 (3 H, s, C-18 Me), 0.90 (3 H, s, C-19 Me), 4.10 (1 H, t, J = 3.0 Hz, H-7 α), 4.65 (1 H, sept, $J_1 = 10.6$ Hz, $J_2 = 4.9$ Hz, H-3 β). Anal. Calcd for C₂₉H₄₈O₅: C, 73.47; H, 10.20. Found: C, 72.84; H, 9.82.

3α-O-Butanoylcholic acid, methyl ester: mp 116-118 °C (from diethyl ether-hexane). ¹H NMR: δ 0.62 (3 H, s, C-18 Me), 0.86 (3 H, s, C-19 Me), 3.84 (1 H, q, J = 2.5 Hz, H-7 β), 3.97 (1 H, t, J = 2.8 Hz, H-12 β), 4.57 (1 H, sept, $J_1 = 10.6$ Hz, $J_2 = 4.9$ Hz, H-3 β). Anal. Calcd for C₂₉H₄₈O₆: C, 71.15; H, 9.88. Found: C, 70.98; H, 9.93.

Preparation of 3β-O-Butanovlrockogenin, Rockogenin (20, 400 mg) was dissolved in 20 mL of anhydrous benzene containing 3 molar equiv of TCEB. Cn.c lipase (1 g) was added and the suspension was shaken at 250 rpm and at 45 °C for 24 h. The enzyme was filtered out, the solvent evaporated, and the crude residue purified by flash chromatography (CHCl₃-AcOEt, 9:0.6), yielding 413 mg (89%) of 3β-O-butanoylrockogenin: mp 195 °C (from MeOH). ¹H NMR: δ 4.68 (1 H, sept, $J_1 = 11$ Hz, $J_2 = 5.7$ Hz, H-3 α). Anal. Calcd for C₃₁H₅₀O₅-MeOH: C, 71.91; H, 10.11. Found: C, 71.06, H, 9.83.

Acknowledgment. We thank the Consiglio nazionale delle Ricerche, Rome, progetto Finalizzato "Biotecnologie e Bioinstrumentazione", and the "Biotecnology Action Programme" of the Commission of the European Communities for financial support of this work.

Conformationally Constrained Peptides. Chirospecific Synthesis of 4-Alkyl-Substituted γ -Lactam-Bridged Dipeptides from L-Aspartic Acid

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Received January 12, 1989

The synthesis of enantiomerically pure γ -lactam-bridged dipeptide analogues of Val-Ala, Ile-Ala, and β -MeLeu-Ala starting from L-aspartic acid is presented. N-(9-Phenylfluorenyl)-L-aspartic acid α -tert-butyl β -methyl diester and N-(9-Phenylfluorenyl)-L-aspartic acid dimethyl ester serve as the educts. They have been successfully alkylated at the β -carbon, C-3, with a variety of electrophiles and with total retention of asymmetric integrity at the α -carbon, followed by a regiospecific reduction of the β -methyl ester. Subsequent oxidation and reductive amination with alanine methyl ester affords the precursors of γ -lactam-bridged dipeptides which have been readily cyclized to the γ -lactams bearing the corresponding value, isoleucine, and leucine side chains.

Introduction

Lactams as conformational constraints in peptide backbones are effective structural tools for probing the active conformations of bioactive peptides.¹⁻⁸ In a number of instances, locking bioactive peptides into active conformers by lactam backbone modification has led to increases in their potency.^{1,9,10} Although several synthetic routes to lactam backbone modified peptides are known,^{1,2,4,5-8,11-17} these methods commonly lack provision

- (2) Kemp, D. S.; Sun, E. T. Tetrahedron Lett. 1982, 23, 3759.
- (3) Kemp, D. S.; McNamara, P. Tetrahedron Lett. 1982, 23, 3761.
 (4) Nagai, U.; Sato, K. Tetrahedron Lett. 1985, 26, 647.
- (5) Thorsett, E. D.; Harris, E. E.; Aster, S.; Peterson, E. R.; Taub, D.;
 Patchett, A. A. Biochem. Biophys. Res. Commun. 1983, 111, 166.
 (6) Manesis, N.; Hassan, M.; Glaser, R.; Goodman, M. Biopolymers
- 1986, 25, S97. (7) Kemp, D. S.; McNamara, P. E. J. Org. Chem. 1985, 50, 5834.

(8) Prochazka, Z.; Lebl, M.; Barth, T.; Hlavacek, J.; Trka, A.; Budesinsky, M.; Jost, K. Collect. Czech. Chem. Commun. 1984, 49, 642.

- (9) Freidinger, R. M.; Perlow, D. S.; Randall, W. C.; Saperstein, R.;
 Arison, B. H.; Veber, D. F. Int. J. Peptide Protein Res. 1984, 23, 142.
 (10) Cascieri, M. A.; Chicchi, G. G.; Freidinger, R. M.; Colton, C. D.;
- Perlow, D. S.; Williams, B.; Curtis, N. R.; McKnight, A. T.; Maguire, J.
 J.; Veber, D. F.; Liang, T. Mol. Pharmacol. 1985, 29, 34.
 (11) Freidinger, R. M.; Perlow, D. S.; Veber, D. F. J. Org. Chem. 1982,
- 47, 104.
- (12) Manesis, N. J.; Goodman, M. J. Org. Chem. 1987, 52, 5331.
 (13) Manesis, N. J.; Goodman, M. J. Org. Chem. 1987, 52, 5342.
 (14) Freidinger, R. M. J. Org. Chem. 1985, 50, 3631.
 (15) Flynn, G. A.; Giroux, E. L.; Dage, R. C. J. Am. Chem. Soc. 1987,

Scheme I. Projections for γ -Lactam-Bridged Dipeptide Analogues



for retention of the amino acid side chain as a substituent on the lactam, place it at C-3 of the lactam, or do not permit continuing extension of the peptide chain. We now present a general synthetic methodology for preparation

⁽¹⁾ Freidinger, R. M.; Veber, D. F.; Perlow, D. S.; Brocks, J. R.; Saperstein, R. Scienc 1980, 210, 656.

^{109, 7914.}

⁽¹⁶⁾ Yanagisawa, H.; Ishihara, S.; Ando, A.; Kanazaki, T.; Miyamoto, S.; Koike, H.; Iijima, Y.; Oizumi, K.; Matsushita, Y.; Hata, T. J. Med. Chem. 1988, 31, 422.

^{(17) (}a) Sham, H. L.; Bolis, G.; Stein, H. H.; Fesik, S. W.; Marcotte,
P. A.; Platner, J. J.; Rempel, C. A.; Greer, J. J. Med. Chem. 1988, 31, 284.
(b) Thaisrivongs, S.; Pals, D. T.; Turner, S. R.; Kroll, L. T. J. Med. Chem.
1988, 31, 1369. (c) Zydowsky, T. M.; Dellaria, J. F., Jr.; Nellans, H. N.
J. Org. Chem. 1988, 53, 5607. (d) Yu, K.-L.; Rajakumar, G.; Srivastava,
M. K. M. K. M. C. M. C. M. M. C. M. C. M. (d) Status and St L. K.; Mishra, R. K.; Johnson, R. L. J. Med. Chem. 1988, 31, 1430.





of enantiomerically pure 4-alkyl-substituted γ -lactambridged dipeptides (1) where the alkyl group at C-4 of the γ -lactam represents the side chain of the former amino acid residue. To demonstrate the versatility of our method, we proposed to synthesize the γ -lactam-bridged analogues of the dipeptides Val-Ala (1A), Ile-Ala (1B), and Leu-Ala (1C) as reflected in Scheme I.

The γ -lactam-bridged dipeptides were to be obtained as projected in sequence 1 from amino acids in a protected form suitable for incorporation into standard peptide synthesis. L-Aspartic acid (2) was chosen as the chiral

$$\underset{H_2N}{\overset{CO_2H}{\longrightarrow}} \xrightarrow{R_3} \underset{O_2R_a}{\overset{CO_2R_g}{\longrightarrow}} \underset{B_1}{\overset{CO_2R_g}{\longrightarrow}} \xrightarrow{R_4} \underset{O_2R_a}{\overset{CH_3}{\longrightarrow}} \xrightarrow{CH_3} (1)$$

educt for the γ -lactam unit because (a) its carbon chain already has all the necessary carbons of the future γ -lactam, (b) it has a β -carboxyl group which allows introduction of an alkyl group via the β -ester enolate, and (c) if differentially esterified, it permits chemical manipulation on each ester separately. The 9-(9-phenylfluorenyl) (PhFl) group¹⁸ was chosen as the N-protecting group. This mode of N-protection insulates the α -center by obstructing removal of the α -proton, thus leading to exclusive formation of the β -ester enolate.^{19,20}

Results and Dicussion

Preparation of γ -Lactam-Bridged Dipeptide Val-Ala (3S, 4S, 2'S)- and (3S, 4R, 2'S)-19. Via the sequence of Scheme II, N-(PhFl)-L-aspartic acid α -tert-butyl β methyl diester (5) was readily prepared from L-aspartic acid (2).²¹ Treating diester 5 with potassium hexamethyldisilazide (KHMDS) in THF at -78 °C followed by trapping the enolate with excess methyl iodide gave alkylated diester 6 in 95% yield as a 2/3, syn/anti²² mixture of diastereoisomers, whose stereochemistry was established subsequently. Similar ratios but lower yields of 6 were obtained with lithium diisopropylamide (LDA) as a base in the presence of HMPT; no reaction was observed when HMPT was absent. Regioselective reduction of the β methyl ester of 6 to alcohol 7 was achieved by treating 6 with DIBAL at -25 °C in THF. The diasteroisomeric alcohols 7a (major isomer, anti) and 7s (minor isomer, syn) were easily separable by chromatography. In contrast to other observations²³ of DIBAL reduction of N-protected aspartic acid diesters, 6 could be reduced in THF without concomitant formation of 4-methylhomoserine lactone. Repeated efforts to reduce diester 6 directly to aldehyde 8 failed; therefore, diastereomerically pure alcohol 7a was oxidized to aldehyde 8a by the dimethyl sulfide-Nchlorosuccinimide procedure.²⁴ Reductive amination²⁵ with L-alanine methyl ester in the presence of sodium cyanoborohydride than gave amination product 9a in 85% yield as a single diastereoisomer.

At this point our synthetic plan was to cleave the tertbutyl ester of 9 to acid 10 and to form the γ -lactam-bridged dipeptide by heating.¹¹ Although many reaction conditions²⁶ were examined, we were unable to find any for hydrolysis of the *tert*-butyl ester without partial cleavage of the PhFl-protected amine. By heating a very dilute solution of free amino acid 12a in DMF in the presence of pyridine, γ -lactam-bridged dipeptide 13 was obtained in very low yield. The major product isolated from this reaction was diketopiperazine 14.

Due to the instability of the N-PhFl group to acidic reaction conditions needed to cleave the tert-butyl ester, the N-protecting group was changed at this stage of the synthesis, where the PhFl group is no longer needed. It had served its function of preventing enolization at the α -carbon of diester 5 and had protected the α -tert-butyl ester in 6 from reduction by DIBAL. Thus alcohol 7a (Scheme III) was first deprotected by hydrogenolysis using Pd/C as catalyst in CH₃OH/HOAc. Reprotection of crude amino alcohol 15a with benzoxycarbonyl chloride (CBZ-Cl) afforded N-CBZ protected amino alcohol 16a in 92% yield. Oxidation²⁴ to aldehyde 17a, followed by reductive amination²⁵ as previously with L-alanine methyl ester produced 18a in yields comparable to those obtained with the N-PhFl-protected compounds. Exposure of aldehyde 17a to silica gel led to slow epimerization at C-3. Therefore, the chromatographic purification of 17a was done rapidly, and traces of the undesired C-3 epimer could be removed at the alkylated amine stage 18a.

Formation of γ -lactam-bridged dipeptide 19a was achieved by first hydrolyzing the tert-butyl ester of 18a in warm formic acid and then neutralizing with pyridine in DMF. γ -Lactam-bridged dipeptide 19a was isolated in 66% yield as a single diastereoisomer. Exactly the same sequence of reactions was used to transform the diastereomeric syn alcohol 7s into the corresponding γ -lactam bridged dipeptide 19s.

Absolute Stereochemistry of N-CBZ γ -Lactam-Bridged Dipeptides 19s and 19a. The initial assignment of absolute stereochemistry was accomplished by ¹H NMR study of the readily available cyclic carbamates 20s,a of the diastereoisomeric alcohols 7 (Scheme III). Alcohol 7, as a pure diastereomer, was hydrogenolyzed using Pd/C as catalyst, and the crude amino alcohol 15 was converted

⁽¹⁸⁾ Bolton, R.; Chapman, N. B.; Shorter, J. J. Chem. Soc. 1964, 1895. Christie, B. D.; Rapoport, H. J. Org. Chem. 1985, 50, 1239.
 Lubell, W. D.; Rapoport, H. J. Am. Chem. Soc. 1987, 109, 236.

⁽²¹⁾ Gmeiner, P.; Feldman, P. L.; Chu-Moyer, M.; Rapoport, H., in preparation.

⁽²²⁾ Following standard usage, the syn and anti designations depend on the relative juxtaposition of the substituent groups when the backbone chain is in its extended form.

⁽²³⁾ Baldwin, J. E.; North, M.; Flinn, A. Tetrahedron Lett. 1987, 28, 3167.

⁽²⁴⁾ Corey, E. J.; Kim, C. U. J. Am. Chem. Soc. 1972, 94, 7586. (25) Borch, R. F.; Bernstein, M. D.; Durst, H. D. J. Am. Chem. Soc.

^{1971, 93, 2897} (26) (a) TFA/CH₂Cl₂, 1/1, at room temperature; (b) TFA at room

temperature; (c) catalytic TsOH·H2O in refluxing benzene; (d) formic acid (95-97%) at room temperature; (e) formic acid (95-97%) at 60 °C; (f) i-PrOH/H₂O/HOAc, 9/9/2, at 95 °C.



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to its cyclic carbamate 20 by reaction with phosgene in toluene in the presence of triethylamine. We considered that 20 should have a chair conformation with the α tert-butyl ester equatorial in both diastereomers and the vicinal methyl group axial in one and equatorial in the other. Analysis by ¹H NMR of cyclic carbamate 20a, derived from the major alcohol 7a, shows two large coupling constants of 7.14 and 8.02 Hz, corresponding to two vicinal axial-H, axial-H couplings, and a small coupling constant of 3.72 Hz corresponding to a vicinal axial-H, equatorial-H coupling. Carbamate 20s, derived from the minor alcohol 7s, shows three small coupling constants of 4.74, 3.52, and 2.8 Hz, corresponding to two vicinal equatorial-H, equatorial-H and one axial-H, equatorial-H coupling. Based on the previous conformational assignment for carbamate 20 and L-aspartic acid as the educt, the major diastereoisomer alcohol is anti. 7a, and has the absolute configuration $2S_{3R}$. The minor alcohol is syn, 7s, and is assigned the configuration 2S.3S.

This assignment of the absolute stereochemistry was confirmed by 2D ¹H NMR NOESY experiments on the diastereoisomeric γ -lactam-bridged dipeptides 19. Major γ -lactam 19a (synthesized from 7a) shows strong dipolar exchange of magnetization (NOE) between protons H₃-H₄, H₄-H_{5 α}, and H_{5 β}-H_{5 α} and a small NOE between H₃-H₄, For the minor γ -lactam 19s (synthesized from 7s) strong NOE's are observed between protons H₄-H_{5 β} and H_{5 β}-H_{5 α} and only very small NOE's between protons H₃-H₄ and H₃-H_{5 α}. These results confirm the assignment of the absolute configuration for alcohols 7s,a based on cyclic carbamates 20. Therefore, dipeptide 19a has the absolute stereochemistry 3S,4S,2'S and 19s is 3S,4R,2'S.

Preparation of N-PhFl γ -Lactam-Bridged Dipeptides Val-Ala (3S,4S,2'S)- and (3S,4R,2'S)-27. Our initial route to the γ -lactam-bridged dipeptide Val-Ala. using N-(PhFl)-L-aspartic acid α -tert-butyl β -methyl diester (5) as a key intermediate, required a detour to change the N-protecting group from PhFl to CBZ due to the instability of the N-PhFl group to acidic hydrolysis conditions needed to cleave the tert-butyl ester. Changing the tert-butyl ester in 9 to a methyl ester should make the γ -lactam available by thermolytic intramolecular condensation with loss of methanol.¹¹ Encouraged by our results in the selective alkylation of diester 5 at C-3 followed by almost total regioselective reduction of the β methyl ester of 6, we applied the same synthetic strategy to N-(PhFl)-L-aspartic acid dimethyl ester (22) as the new intermediate (Scheme IV).

L-Aspartic acid dimethyl ester hydrochloride (21) was obtained from L-aspartic acid in 98% yield by esterification with methanol/thionyl chloride. N-Alkylation with 9-

Scheme IV. Synthesis via Dimethyl Aspartate



bromo-9-phenylfluorene proceeded in 90% yield to N-(PhFl)-L-aspartic acid dimethyl ester (22).²¹ Then C-alkylation of dimethyl ester 22 with CH₃I was carried out under identical conditions as in the previous alkylation of α -tert-butyl β -methyl diester 5, producing 23 in 82% yield as a 3/2, syn/anti (see below) mixture of diastereoisomers together with 15% of the easily separated C-3 dialkylated product. Reduction of mixture 23 with DIBAL in THF at -35 °C occurred highly regiospecifically to give mixed alcohols 24 in 93% yield without any formation of 4methylhomoserine lactone.²³ Oxidation of the 3/2 mixture of alcohols 24 as previously described gave 78% of diastereoisomeric aldehydes 25, easily separable by MPLC. Reductive amination of major aldehvde 25s (pure syn diastereoisomer) with L-alanine methyl ester using NaC-NBH₃ as the reducing agent afforded a 95% yield of amination product 26s. Conversion of 26s to γ -lactam 27s was best achieved by refluxing in toluene for 20 h. The absolute configuration of the diastereoisometric γ -lactams 27a,s could not be assigned unambiguously by ¹H NMR spectroscopy, due to overlapping chemical shifts of protons HC-3 and H_2C-5 and the small differences between the coupling constants of these protons in 27a and 27s.

The absolute configuration of the diastereoisomeric γ -lactams 27a,s could be assigned, however (Scheme IV), by hydrogenolytic removal of the N-protecting group of N-PhFl γ -lactam 27a and N-CBZ γ -lactam 19a, the stereochemistry of the latter being known. Comparison of their ¹H NMR spectra showed them to be identical. Thus the absolute stereochemistry can be assigned at 3S, 4S, 2'S

Table I. ¹H NMR Data for γ -Lactam-Bridged Dipeptides

	compound (3 <i>S</i> ,4 <i>R</i> ,2' <i>S</i>)-19	ð, ppm (J, Hz)								
		CH ₃ C-4	CH ₃ C-2′ 1.41	HC-4 2.27	HC-5		CO ₂ CH ₃	HC-3	HC-2′	
_		1.25			2.95	3.50	3.71	4.08	4.85	
		(d, 6.4)	(d, 7.5)	(m)	(t, 9.3)	(dd, 9.2, 8.1)		(t, 8.8)	(q, 7.5)	
	(3S.4S.2'S)-19	0.98	1.42	2.88	3.08	3.54	3.72	4.42	4.84	
	(, , , , , ,	(d. 7.0)	(d. 7.5)	(m)	(d, 9.6)	(dd, 9.5, 5.9)		(t, 5.8)	(q, 7.5)	
	(3S.4R.2'S)-27	0.17	1.27	2.07	2.52	3.32	3.72	2.50	4.75	
	(,,,	(d. 6.7)	(d. 7.3)	(m)	(dd, 9.3, 8.4)	(dd, 9.3, 8.3)		(d, 8.6)	(q, 7.4)	
	(3S.4S.2'S)-27	1.09	1.23	1.50	2.77	3.00	3.69	2.75	4.73	
	(,,,,,	(d. 7.0)	(d. 7.4)	(m)	(d. 9.4)	(dd. 9.4, 5.4)		(d, 6.7)	(g, 7.4)	
	(3S.4R.2'R)-27	0.13	1.37	1.93	2.62	3.29	3.61	2.58	4.83	
	(00),,,	(d. 6.7)	(d. 7.4)	(m)	(dd. 8.4, 7.8)	(t, 8.8)		(d, 7.8)	(q, 7.4)	
	(3S 4S.2'R)-27	0.99	1.30	1.48	2.69	3.11	3.58	2.79	4.80	
	(02):-;=:;/=:	(d. 7.0)	(d, 7.4)	(m)	(d. 9.7)	(dd, 9.7, 5.2)		(d. 6.7)	(q, 7.4)	
	(3S, 4R, 2'S)-32	(4, 110)	1.29	1.93	2.61	3.37	3.73	2.59	4.76	
	(00,110,20) 02		(d. 7.4)	(m)	(dd. 9.5, 7.6)	(t, 8.8)		(d. 8.5)	(a. 7.4)	
	(3S 4R 2'S)-39		1.42	2.07-2.15	3.04	3.44	3.72	4.29	4.87	
	(00, 11, 20) -00		(d, 7.5)	(m)	(t, 9.2)	(t, 9.0)		(dd, 9.9, 9.4)	(q, 7.5)	

for 27a and 3S,4R,2'S for 27s.

Optical Purity of 27. To test if steric integrity had been lost at any of the chiral centers in 27 during its preparation or chromatographic purification, one enantiomer of each diastereomeric pair of 27 was synthesized. HPLC analysis of (3S,4S,2'S)-, (3S,4R,2'S)-27 and (3S,4S,2'R)-, (3S,4R,2'R)-27, obtained from aldehydes 25s and 25a, respectively, by reductive amination with D-alanine methyl ester proved that the N-PhFl-protected γ lactam-bridged dipeptides Val-Ala (27) are >99.5% enantiomerically pure. This establishes that the N-CBZprotected γ -lactam-bridged dipeptides 19 also are >99.5% enantiomerically pure, since N-deprotection of 27 and 19 give identical products.

Preparation of N-PhFl-Protected γ -Lactam-Bridged Dipeptide Ile-Ala (3S,4R,2'S)-32. Extension of our synthetic methodology to the preparation of the N-PhFl-protected γ -lactam-bridged dipeptide Ile-Ala (3S,4R,2'S)-32, starting from N-(PhFl)-L-aspartic acid dimethyl ester (22), is shown in Scheme V. Alkylation of 22 as previously but using ethyl iodide led to dialkylation at C-3, and 28 was isolated as the minor reaction product in low yields. Changing the electrophile to ethyl triflate²⁷ resulted in a dramatic decrease in reaction time, and 28 was isolated in 80% yield as a 3.5/1 mixture of syn/anti diastereoisomers, separable by MPLC; dialkylation at C-3 of 22 was observed only to the extent of 5%. The regioselective reduction of 28s (syn, major isomer) using DIBAL in THF failed. Instead of alcohol 29s, 4ethylhomoserine lactone was isolated as the major product.²³ No lactone formation had been observed in the DIBAL reduction of ester 23 to alcohol 24; therefore, lactone formation is strongly dependent on the size of the alkyl group at C-3. Lactone formation could be minimized, however, by conducting the DIBAL reduction in toluene at -50 °C for 20 min and oxidizing crude alcohol 29s directly to aldehyde 30s in 59% yield from diester 28. Reductive amination with L-alanine methyl ester and NaC-NBH₃ afforded a 64% yield of amination product 31s. Condensation to γ -lactam 32s was then achieved in 86% yield by refluxing 31 for 3 h in p-xylene.

Assignment of the absolute configuration at C-4 of γ lactam **32s** was made by correlation of its ¹H NMR spectrum with those for the C-4 epimeric γ -lactams **27s,a**. Pertinent chemical shifts and coupling constants are collected in Table I. The close coincidence of the coupling constants for HC-3 and H₂C-5 of **32s** with those of



Scheme V. Synthesis of Ile-Ala Analogue



(3S,4R,2'S)-27, and the fact that the protons of the C-4 alkyl group show the same upfield shift due to an anisotropic shielding effect of the N-PhFl group, allow assignment of 3S,4R,2'S as the configuration of 32s (Table I).

Preparation of N-CBZ γ -Lactam-Bridged Dipeptide β -MeLeu-Ala (3S,4R,2'S)-39. As demonstrated in the previous syntheses, γ -lactam-bridged dipeptide Val-Ala can be prepared either from N-(PhFl)-L-aspartic acid α tert-butyl β -methyl diester (5) to give the N-CBZ γ -lactams 19 (Scheme III) or, in fewer steps, from N-(PhFl)-L-aspartic acid dimethyl ester (22) to give N-PhFl γ -lactams 27 (Scheme IV). Although dimethyl ester 22 is a versatile educt for the synthesis of N-PhFl γ -lactam-bridged dipeptide Ile-Ala 32s (Scheme V), its synthetic utility is somewhat diminished because of the instability of alcohol 29 toward lactone formation. Anticipating similar problems during the preparation of γ -lactam-bridged dipeptide β -MeLeu-Ala 39, we chose N-(PhFl)-L-aspartic acid α tert-butyl β -methyl diester (5) as the educt and, at the same time, demonstrated another synthetic application of 5 (Scheme VI).

The alkylation of 5 was carried out using 200 mol % of KHMDS in THF at -78 °C and trapping the enolate with 200 mol % of isopropyl triflate, added as a freshly prepared 0.6 M solution in hexane.²⁷ The process of deprotonation with KHMDS and trapping the enolate with isopropyl triflate was repeated twice. After the reaction was quenched with 1 M H_3PO_4 , 33 was isolated in 43% yield

Table II. ¹H NMR Data for 2-Amino-4-hydroxy-3-methylbutanoates and β -Substituted Aspartates

compound	CH ₃ C-3	HC-1′	CH ₃ C-1'	HC-2	HC-3	HC-4	$\rm CO_2Bu^t$	CO ₂ CH ₃
(2S,3S)-7	0.86			2.70	1.65	3.40	1.17	
	(d, 7.2)			(s)	(m)	(dd, 11, 3.2)		
						3.51		
						(t, 11)		
(2S, 3R)-7	0.65			2.45	1.85	3.40	1.16	
	(d, 7.0)			(d, 8.3)	(m)	(dd, 11, 8.5)		
						3.54		
						(dd, 11, 3.3)		
(2S, 3S) - 16	0.65			4.59	2.35	3.23	1.47	
	(d, 6.9)			(dd, 7.8, 2.7)	(m)	(m)		
						3.45		
					0.00	(m)		
(2S, 3R) - 16	1.01			4.26	2.06	3.52	1.47	
	(a, 7.0)			(aa, 8.4, 6.3)	(m)	(d, 10.5)		
						3.07 /33 11 5 9 0)		
(15 25) 28		1 56_1 70	0.77	2 70	9.97	(dd, 11.5, 2.9)		9.94
(20,00)-20		(m)	(+ 7.4)	2.75 (dd 10.2 66)	2.37 (m)			0.24 9.51
(25,3R)-28		1 14	(1, 7.4)	(uu, 10.2, 0.0) 9.85	9 47			3.01
(20,011)-20		(m)	(t, 7, 4)	(dd 114 88)	2.47 (m)			3.76
		1.41	(0, 1.4)	(44, 11.4, 0.0)	(111)			0.70
		(m)						
(2S.3S)-33		1.96	0.41	2.66	2.14		1.24	3.68
(,-,,		(m)	(d, 6.5)	(d, 4.9)	(dd, 10, 4.9)			
			0.67					
			(d, 6.5)					
(2S, 3R)-33		2.05	0.54	2.90	2.24		1.10	3.71
		(m)	(d, 6.6)	(d, 5.3)	(dd, 8.0, 6.0)			
			0.83					
			(d. 6.6)					

Scheme VI. Synthesis of Leu-Ala Analogue



as a 5/1, syn/anti, mixture of diastereoisomers, separable by MPLC, together with 45% of recovered diester 5. Reduction of major isomer 33s to alcohol 34s was done as previously described in the reduction of 28. Alcohol 34s was isolated in 78% yield and showed no tendency to form lactone even under chromatographic purification. Hydrogenolytic (H₂, Pd/C) deprotection of alcohol 34s in CH₃OH/HOAc followed by reprotection afforded N-CBZ alcohol 35s in 66% yield together with 30% of 4-isopropylhomoserine lactone 36s. Acid-catalyzed lactone formation occurred during the hydrogenolysis of 34s and can be minimized by shortening the reaction time and omitting the acetic acid. Oxidation to aldehyde 37s (80% yield) and reductive amination to 38s (52% yield) with L-alanine methyl ester was carried out in the same way as described earlier for compounds 17 and 18. Heating 38s neat to 60 °C led to slow conversion to γ -lactam 39s, which

was prepared in 62% vield analogously to the previously described preparation of γ -lactam 19. Assignment of the absolute configuration at C-4 of γ -lactam 39s was made by a 2D ¹H NMR NOESY analysis. γ -Lactam 39s shows a strong NOE only between protons $H_4-H_{5\beta}$ and $H_{5\beta}-H_{5\alpha}$ and a weak NOE between protons H_3-H_4 and $H_3-H_{5\alpha}$. Therefore the absolute configuration of γ -lactam 39s is 3S,4R,2'S (Table I).

Synthesis of 3-alkylaspartic acids has been reported,^{28,29} and recently 3-methylaspartic acid was found in the cyclic peptide toxin cyanogenosin RR, isolated from Microcystis aeruginosa.³⁰ The first synthesis²⁸ proceeded from ditert-butyl N-formylaspartate and gave a mixture of α - and β -alkylated products. For the β -diastereomer, erythro configuration was assigned by analogy³¹ and absolute stereochemistry derived by conversion to 3-methylaspartic acid and comparison of specific rotations.³² The second synthesis²⁹ involves incubation of 3-alkylfumaric acid with the enzyme 3-methylaspartate ammonia-lyase (EC 4.3.1.2) for several days. The overall yield of purified 3-alkylaspartic acid varies from 20 to 40% depending on the alkyl group, and in the case of n-butyl, no amination occurred. The relative stereochemistry of 3-methyl- and 3-ethylaspartic acid has been assigned from the coupling constants of HC-2 and HC-3 of their N-(trifluoroacetyl)aspartic anhydrides. The absolute configuration at C-3 was assigned by degrading 3-ethylaspartic acid to 3-ethylsuccinic acid for which specific rotations have been reported.³³⁻³⁵ The

(31) Seebach, D.; Wasmuth, D. Helv. Chim. Acta 1980, 63, 197. (32) Bodansky, M.; Marconi, G. G. J. Antibiot. 1970, 23, 238.

(34) Fredga, A. Arkiv Kemi 1947, 24A, 1.

1.

(35) Berner, E.; Leonardsen, R. Justus Liebigs Ann. Chem. 1939, 538,

⁽²⁸⁾ Seebach, D.; Wasmuth, D. Angew. Chem., Int. Ed. Engl. 1981, 20, 971.

⁽²⁹⁾ Akhtar, M.; Botting, N. P.; Cohen, M. A.; Gani, D. Tetrahedron 1987, 43, 5899. (30) Painuly, P.; Perez, R.; Fukai, T.; Shimizu, Y. Tetrahedron Lett.

^{1988, 29, 11.}

⁽³³⁾ Wren, H.; Crawford, J. J. Chem. Soc. 1937, 230.

Conformationally Constrained Peptides

correlation between the specific rotation and the absolute configuration was established by X-ray analysis of an ergoflavine derivative which gave (-)-methylsuccinic acid on oxidation.³⁶ The synthetic methodology described in the present work not only has the advantage of affording a variety of 3-alkylaspartic acids in high yields and in a fully protected form suitable for peptide synthesis, but also allows the unambiguous determination of their absolute stereochemistry by definitive NMR experiments. In Table II, the NMR data for the diastereoisomeric N-PhFl-protected 3-ethyl- and 3-isopropylaspartic acid diesters 28s,a and 33s,a are summarized together with the data for the diastereoisomeric N-PhFl- and N-CBZ-2-amino-4hydroxy-3-methylbutanoic acid *tert*-butyl esters 7s and 7a and 16s and 16a.

Summary

The protected γ -lactam-bridged dipeptides Val-Ala, Ile-Ala, and β -MeLeu-Ala have been prepared in enantiomerically pure form starting from N-(PhFl)-L-aspartic acid α -tert-butyl β -methyl diester (5) or N-(PhFl)-L-aspartic acid dimethyl ester (22), both readily available from L-aspartic acid (2).²¹ The N-PhFl protecting group directs regiospecific alkylation of 5 and 22 to C-3, without any loss of steric integrity at the α -chiral center. Furthermore, the bulk of the PhFl group contributed to the specific reduction of the β -methyl ester in 6, 23, 28, and 33 without reducing the α -methyl or α -tert-butyl ester.

The synthetic routes described in this paper represent efficient and general methodology for the synthesis of enantiomerically pure 4-alkyl γ -lactam-bridged dipeptides. Such peptides then can be incorporated into polypeptides to induce a degree of conformational constraint.

Experimental Section

General. Tetrahydrofuran (THF) was distilled from sodium/benzophenone immediately before use; chloroform was distilled from P₂O₅; methylene chloride, acetonitrile, isopropyl alcohol, triethylamine, pyridine, and trimethylsilyl chloride were distilled from CaH₂; toluene and hexane were distilled from NaH; methanol from magnesium; and dimethylsulfide (Me₂S) from sodium. N-Chlorosuccinimide (NCS) was crystallized from benzene. Column chromatography was performed with 230-400 (low-pressure chromatography) and 70-230 (gravity chromatography) mesh silica gel. Preparative medium-pressure liquid chromatography (MPLC) was performed with glass columns and 230-400 mesh silica gel. High-pressure liquid chromatography (HPLC) was done on a 4.6 \times 250 mm, 5 μ m LiChrosorb Si 60 normal-phase silica gel column at a flow rate of 1 mL/min. Thin-layer chromatography (TLC) was done on silica 60/F-254 aluminum-backed plates (E. Merck). ¹H NMR spectra were recorded at 250 MHz or 500 MHz in CDCl₃ unless otherwise noted. Chemical shifts are reported in ppm (δ units) downfield from internal tetramethylsilane or internal sodium 3-(trimethylsilyl)propionate- d_4 (TSP) for spectra taken in D₂O. Melting points were determined on a Büchi melting point apparatus and are uncorrected. Elemental analyses were performed by the Analytical Laboratory, College of Chemistry, University of California, Berkeley. Unless otherwise noted, reactions were conducted under a nitrogen atmosphere. Final product solutions were dried over $MgSO_4$ or Na_2SO_4 , filtered, and evaporated under reduced pressure on a Berkelev rotary evaporator. In cases where identical reactions have been carried out with diastereomeric educts, only the experimental procedure for the major diastereomer is described.

N-(9-Phenylfluorenyl)-L-aspartic acid α -tert-butyl β methyl diester (5) was prepared from L-aspartic acid via the β -methyl ester 3 and its N-(9-phenylfluorenyl) derivative 4 as described.²¹

N-(9-Phenylfluorenyl)-3-methyl-L-aspartic Acid α -tert-Butyl β -Methyl Diester (6). To a stirred solution of KHMDS (5.6 mL, 3.35 mmol, 0.6 M in toluene) in 50 mL of THF was added 5 (1.06 g, 2.4 mmol) dissolved in 10 mL THF at -78 °C dropwise. The pale vellow solution was stirred at -78 °C for 45 min, CH₃I (0.46 mL, 7.2 mmol) was added, and the resulting solution was stirred at -78 °C for 1 h. The reaction was quenched with 1 mL of MeOH at -78 °C, warmed up to room temperature, and partioned between 80 mL of 1 M H_3PO_4 and 80 mL of Et_2O . The organic layer was washed with 50 mL of H₂O and 50 mL of saturated $Na_2S_2O_3$, dried, and evaporated. The oily residue was chromatographed (low pressure, hexane/EtOAc, 12/1) to give 6 as a thick oil in 95% yield and a 2/3 ratio of syn/anti diastereoisomers according to ¹H NMR: ¹H NMR δ 0.96 (d, J = 7.0, CH₂C-3), 1.04 (d, J = 7.4, CH₃C-3), 1.08, 1.12 (2 s, CO₂Bu^t), 2.47 (m, HC-3), 2.86 (m, HC-2), 3.40, 3.53 (2 s, CO₂CH₃), 7.11-7.33 (m, 11 H, Ar), 7.58-7.66 (m, 2 H, Ar). Anal. Calcd for C₂₉H₃₁NO₄: C, 76.1; H, 6.8; N, 3.1. Found: C, 76.2; H, 6.9; N, 3.1.

tert -Butyl (2S,3S)-2-[(9-Phenylfluorenyl)amino]-3methyl-4-hydroxybutanoate [(2S,3S)-7] and tert-Butyl (2S,3R)-2-[(9-Phenylfluorenyl)amino]-3-methyl-4hydroxybutanoate [(2S,3R)-7]. Into a solution of 6 (1.02 g, 2.23 mmol, mixture of diastereomers) in 75 mL of THF was dropped DIBAL (6.9 mL, 6.9 mmol, 1 M in hexane) at -30 °C. The clear solution was stirred at -30 °C for 4 h, 0.33 mL of acetone and, after 10 min, 2 mL of MeOH were added, and the reaction mixture was partitioned between 75 mL of 1 M H₃PO₄ and 150 mL of Et₂O. Saturated NaHCO₃ (50 mL) was added, the mixture was filtered, and the organic phase was washed with brine, dried, and evaporated. The residue was chromatographed (low pressure, hexane/EtOAc, 5/1) to give 350 mg of (2S,3S)-7 and 460 mg of (2S,3R)-7 (85% yield).

(2S, 3R)-7: ¹H NMR δ 0.65 (d, 3 H, J = 7.0, CH₃C-3), 1.16 (s, 9 H, CO₂Bu^t), 1.85 (m, 1 H, HC-3), 2.45 (d, 1 H, J = 8.3, HC-2), 3.40 (dd, 1 H, J = 11, 8.5, HC-4), 3.54 (dd, 1 H, J = 11, 3.3, HC-4), 7.15–7.47 (m, 11 H, Ar), 7.68–7.71 (m, 2 H, Ar). Anal. Calcd for C₂₈H₃₁NO₃: C, 78.3; H, 7.3; N, 3.3. Found: C, 78.4; H, 7.2; N, 3.3.

(2S,3S)-7: ¹H NMR δ 0.86 (d, 3 H, J = 7.2, CH₃C-3), 1.17 (s, 9 H, CO₂Bu^t), 1.65 (m, 1 H, HC-3), 2.70 (s, 1 H, HC-2), 3.0 and 3.3 (2 s, br, OH, NH), 3.40 (dd, 1 H, J = 11, 3.2, HC-4), 3.51 (t, 1 H, J = 11, HC-4), 7.19–7.53 (m, 11 H, Ar), 7.67–7.71 (m, 2 H, Ar).

tert-Butyl (2S,3R)-2-[(9-Phenylfluorenyl)amino]-3methyl-4-oxobutanoate [(2S, 3R)-8]. To a stirred suspension of N-chlorosuccinimide (560 mg, 4.2 mmol) in 10 mL of toluene was added (CH₃)₂S (0.42 mL, 5.7 mmol) at 0 °C. The suspension was cooled to -25 °C, and a solution of (2S,3R)-7 (450 mg, 1.05 mmol) in 4 mL of toluene was added dropwise. The reaction mixture was stirred for 4 h at -25 °C, and then Et₃N (0.56 mL, 4.2 mmol) in 1 mL of toluene was added. The cooling bath was removed and after 5 min 25 mL of Et₂O was added, and the mixture was washed with 0.5 M H₃PO₄ (25 mL) and H₂O (25 mL), dried, and evaporated. The oily residue was chromatographed (low pressure, hexane/EtOAc, 10/1) to leave (2S, 3R)-8 as a thick oil (360 mg, 80%): ¹H NMR δ 0.95 (d, 3 H, J = 6.9, CH₃C-3), 1.17 (s, 9 H, CO_2Bu^t), 2.33 (m, 1 H, HC-3), 2.94 (t, 1 H, J = 6.7, HC-2), 3.28 (s, 1 H, J = 7.5, NH), 7.17–7.42 (m, 11 H, Ar), 7.67–7.73 (m, 2 H, Ar), 9.37 (d, 1 H, J = 1.5, CHO).

(2S, 3S)-8: ¹H NMR δ 1.01 (d, 3 H, J = 6.9, CH₃C-3), 1.19 (s, 9 H, CO₂Bu^t), 2.35 (m, 1 H, HC-3), 3.13 (m, 2 H, HC-2, NH), 7.17-7.40 (m, 11 H, Ar), 7.66-7.71 (m, 2 H, Ar), 9.14 (s, 1 H, CHO).

Reductive Amination Product. (2S, 3S, 2'S)-9. A solution of (2S, 3R)-8 (700 mg, 1.64 mmol), L-alanine methyl ester hydrochloride (1.12 g, 8.2 mmol), and NaCNBH₃ (82 mg, 1.3 mmol) in 50 mL of MeOH was stirred at room temperature for 45 min. The reaction mixture was evaporated, and the oily residue was partitioned between 50 mL of saturated NaHCO₃ and 100 mL of Et₂O. The H₂O layer was extracted with 40 mL of Et₂O, and the combined organic phase was washed with 30 mL of H₂O, dried, and evaporated. The residue was chromatographed (low pressure, hexane/EtOAc, 4/1) to leave 642 mg (76% yield) of (2S,3S,2'S)-9 as a thick oil: ¹H NMR δ 0.80 (d, 3 H, J = 6.9, CH₃C-3), 1.16 (s, 9 H, CO₂Bu¹), 1.19 (d, 3 H, J = 6.9, CH₃C-2'), 1.71 (m, 1 H, HC-3), 2.36 (dd, 1 H, J = 11.7, 7.4, HC-4), 2.46 (dd, 1 H, J = 11.7, 6.3, HC-4), 2.48 (d, 1 H, J = 5.0, HC-2), 3.15 (q, 1 H, J = 6.9, HC-2'),

⁽³⁶⁾ McPhail, A. T.; Sim, G. A.; Asher, J. D. M.; Robertson, J. M.; Silverton, J. V. J. Chem. Soc. B 1966, 18.

3.70 (s, 3 H, CO₂CH₃), 7.18–7.41 (m, 11 H, Ar), 7.64–7.67 (m, 2 H, Ar).

(2S, 3R, 2'S)-9: ¹H NMR δ 0.84 (d, 3 H, J = 6.9, CH₃C-3), 1.15 (d, 3 H, J = 7.0, CH₃C-2'), 1.17 (s, 9 H, CO₂Bu^t), 1.60 (m, 1 H, HC-3), 2.27 (dd, 1 H, J = 12, 7.2, HC-4), 2.42 (dd, 1 H, J = 12, 6.7, HC-4), 2.51 (s, br, 1 H, HC-2), 3.05 (s, br, 1 H, NH), 3.21 (q, 1 H, J = 7.0, HC-2'), 3.70 (s, 3 H, CO₂CH₃), 7.18–7.49 (m, 1 H, Ar), 7.64–7.73 (m, 2 H, Ar).

(2S,3S,2'S)-11. A solution of (2S,3S,2'S)-9 (600 mg, 1.17 mmol) and Pd/C (10%, 300 mg) in 50 mL of MeOH/HOAc, 20/1, was shaken under 50 psi of H₂ for 16 h. The reaction mixture was filtered, the filtrate was evaporated, and the residue was partitioned between 30 mL of H₂O and 30 mL of Et₂O. The H₂O layer was basified with saturated NaHCO₃ and extracted with CHCl₃ (3 × 30 mL), and the organic layer was dried and evaporated to leave (2S,3S,2'S)-11 as a colorless oil in 96% yield: ¹H NMR δ 0.96 (d, 3 H, J = 7.0, CH₃C-3), 1.27 (d, 3 H, J = 7.1, CH₃C-2'), 1.47 (s, 9 H, CO₂Bu⁺), 2.02 (m, 1 H, HC-3), 2.53 (dd, 2 H, J = 7.0, 3.5, H₂C-4), 3.29 (m, 2 H, HC-2, HC-2'), 3.71 (s, 3 H, CO₂CH₃).

Deprotected γ -Lactam Dipeptide (3S,4S,2'S)-13. From (3S,4S,2'S)-27. A solution of 30 mg (0.07 mmol) of (3S,4S,2'S)-27 and 20 mg of Pd/C (10%) in 8 mL of MeOH/HOAc (20:1) was mechanically shaken under 50 psi of H₂ for 16 h. The reaction mixture was filtered through Celite and evaporated, the residue was partitioned between 10 mL of H_2O and 10 mL of Et_2O , and the water layer was basified with saturated NaHCO3 and extracted with $CHCl_3$ (3 × 10 mL). The combined organic layer was dried and evaporated to leave 12 mg (88%) of (3S,4S,2'S)-13 as a pale yellow oil. From (3S,4S,2'S)-19. A suspension of 15 mg of (3S,4S,2'S)-19 and 4 mg of Pd/C (10%) in 2 mL of EtOAc was hydrogenolyzed with H_2 at 1 atm for 30 min. The reaction mixture was filtered through Celite and evaporated to give 5 mg of (3S,4S,2'S)-13 as a pale yellow oil: ¹H NMR δ 1.08 (d, 3 H, J = 7.1, CH₃C-4), 1.41 (d, 3 H, J = 7.6, CH₃C-2'), 2.61 (m, 1 H, HC-4), 3.04 (dd, 1 H, J = 9.4, 1.8, HC-5), 3.47 (dd, 1 H, J = 9.4, 6.1, HC-5), $3.60 (d, 1 H, J = 7.4, HC-3), 3.73 (s, 3 H, CO_2CH_3), 4.85 (q, 1 H, J)$ J = 7.5, HC-2').

Diketopiperazine 14. A solution of (2S,3S,2'S)-11 (154 mg, 0.56 mmol) in 4 mL of formic acid (95-97%) was heated to 75 °C for 3.5 h. After 3.5 h, no 11 could be detected by TLC (silica gel, CHCl₃/MeOH/pyridine, 10:1:0.5), and the reaction mixture was evaporated and dried (Kugelrohr, 55 °C, 0.1 mmHg, over night), to leave crude (2S, 3S, 2'S)-12 as a thick syrup. A heterogeneous mixture of 125 mg (0.404 mmol) of crude (2S,3S,2'S)-12 and 65 μ L (0.808 mmol) of pyridine in 4 mL of cold DMF was added slowly (syringe pump, 1 h) to 150 mL of DMF at 75 °C. After complete addition, the homogeneous solution was stirred at 75 °C for 3 h, the reaction mixture was evaporated, and the residue was partitioned between 40 mL of CHCl₃ and 30 mL of saturated NaHCO₃. The organic layer was dried and evaporated to leave 60 mg (75%) of a mixture of 14 and (3S,4S,2'S)-13 (95/5, according to ¹H NMR): ¹H NMR δ 0.98 (d, 3 H, J = 7.1, CH₃C-3), 1.44 (d, 3 H, J = 7.5, CH_3C-2'), 2.94 (m, 1 H, HC-3), 3.12 (d, 1 H, J = 9.7, HC-4), 3.58 (dd, 1 H, J = 9.7, 5.7, HC-4), 3.74 (s, 3 H, CO_2CH_3), 4.62 (t, 1 H, J = 6.1, HC-2), 4.84 (q, 1 H, J = 7.5, HC-2'), 6.58 (s, br, 1 H, CONH), 8.32 (d, 1 H, J = 1, NH-O).

tert-Butyl (2S,3R)-2-Amino-3-methyl-4-hydroxybutanoate [(2S,3R)-15]. A solution of (2S,3R)-7 (180 mg, 0.42 mmol) and 90 mg of Pd/C (10%) in 12 mL of MeOH/HOAc (20/1) was mechanically shaken under 50 psi of H₂ for 16 h. The reaction mixture was filtered through Celite and evaporated, and the residue was partitioned between 20 mL of H₂O and 20 mL of Et₂O. The water layer was basified with saturated NaHCO₃ and extracted with CHCl₃ (3 × 20 mL). The organic layer was dried and evaporated to leave (2S,3R)-15 as a colorless oil in quantitative yield: ¹H NMR δ 0.90 (d, 3 H, J = 7.0, CH₃C-3), 1.48 (s, 9 H, CO₂Bu⁴), 1.98 (m, 1 H, HC-3), 2.83 (s, br, NH₂, OH), 3.27 (d, 1 H, J = 7.9, HC-2), 3.64 (m, 2 H, H₂C-4).

(2S, 3S)-15: ¹H NMR δ 0.94 (d, ³ H, J = 7.2, CH₃C-3), 1.47 (s, 9 H, CO₂Bu^t), 2.14 (m, 1 H, HC-3), 2.48 (s, br, NH₂, OH), 3.62 (dd, 1 H, J = 10.9, 4.9, HC-4 and d, 1 H, J = 3.1, HC-2), 3.90 (dd, 1 H, J = 10.9, 3.0, HC-4).

tert -Butyl (2S,3R)-2-[(Benzoxycarbonyl)amino]-3methyl-4-hydroxybutanoate [(2S,3R)-16]. To a stirred solution of (2S,3R)-15 (132 mg, 0.7 mmol) and pyridine (85 μ L, 1.05 mmol) in 5 mL of dry CH₃CN was added CBZ-Cl (120 µL, 0.84 mmol) dissolved in 0.5 mL of CH₃CN drop-by-drop at 0 °C under N₂. The mixture was stirred for 2 h at 0 °C, and then again pyridine (85 μ L, 1.05 mmol) followed by CBZ-Cl (120 μ L, 0.84 mmol) in 0.5 mL CH₃CN were added. Stirring was continued for 1 h, the reaction mixture was evaporated, and the residue was partitioned between 25 mL of 1 M H_3PO_4 and 25 mL of CHCl₃. The H_2O layer was extracted with CHCl_3 (2 \times 20 mL), and the combined organic layer was dried and evaporated. The crude product was purified by chromatography (low pressure, silica gel, hexane/ EtOAc, 2/1) to leave (2S, 3R)-16 (230 mg, 92%) as a pale yellow oil: ¹H NMR δ 1.01 (d, 3 H, J = 7.0, CH₃C-3), 1.47 (s, 9 H, CO₂Bu^t), 2.06 (m, 1 H, CH-3), 2.45 (s, br, 1 H, OH), 3.52 (d, br, 1 H, J = 10.5, HC-4, 3.67 (dd, 1 H, J = 11.5, 2.9, HC-4, 4.26 (dd, 1 H, J = 11.5, 2.9, 1.5) (dd, 1 H, J = 11.5, 2.9, 1.5) (dd, 1 H, J = 11.5, 2.9, 1.5) (dd, 1 H, J = 11.5, 2.5) (dd, 1 H, J = 11.5, 2.5) (dd, 1 H, J = 11.5, 2.5) (dd, 1 H, J = 11.51 H, J = 8.4, 6.3, HC-2, 5.11 (s, 2 H, CH₂Ar), 5.76 (d, 1 H, J =8.4, NH), 7.31-7.37 (s, 5 H, Ar). Anal. Calcd for C₁₇H₂₅NO₅: C, 63.1; H, 7.8; N, 4.3. Found: C, 63.1; H, 7.9; N, 4.3.

(25,35)-16: ¹H NMR δ 0.65 (d, 3 H, J = 6.9, CH₃C-3), 1.47 (s, 9 H, CO₂Bu^t), 2.35 (m, 1 H, HC-3), 3.23 (m, 1 H, HC-4), 3.45 (m, 1 H, HC-4), 4.0 (s, br, 1 H, OH), 4.59 (dd, 1 H, J = 7.8, 2.7, HC-2), 5.12 (s, 2 H, CH₂Ar), 5.58 (d, 1 H, J = 7.3, NH), 7.34–7.37 (s, 5 H, Ar).

tert-Butyl (2S,3R)-2-[(Benzoxycarbonyl)amino]-3methyl-4-oxobutanoate [(2S, 3R)-17]. To a stirred suspension of NCS (314 mg, 2.35 mmol) in 10 mL of dry toluene was added Me_2S (235 $\mu L,$ 3.21 mmol) at 0 °C under $N_2.\,$ The suspension was cooled to -25 °C, and (2S,3R)-16 (190 mg, 0.59 mmol) in 2 mL of toluene was added drop-by-drop. Stirring was continued for 4 h at -25 °C, Et₃N (327 µL, 2.35 mmol) in 0.5 mL of toluene was added, the cold bath was removed, and after 10 min, the reaction mixture was partitioned between 25 mL of 1 M H₃PO₄ and 30 mL of Et_2O . The organic layer was washed with 20 mL of H_2O , dried, and evaporated. The oily residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 5/1) to leave (2S,3R)-17 in 75% yield as a colorless oil: ¹H NMR δ 1.12 (d, 3 H, J = 7.2, CH₃C-3), 1.42 (s, 9 H, CO₂Bu^t), 3.09 (m, 1 H, HC-3), 4.69 (dd, 1 H, J = 7.9, 3.7, HC-2), 5.13 (s, 2 H, CH₂Ar), 5.57 (d, 1 H, J = 8.0, NH, 7.36 (s, 5 H, Ar), 9.69 (s, 1 H, CHO). Anal. Calcd for C₁₇H₂₃NO₅: C, 63.5; H, 7.2; N, 4.4. Found: C, 63.5; H, 7.3; N, 4.3.

(2S,3S)-17: ¹H NMR δ 1.08 (d, 3 H, J = 7.1, CH₃C-3), 1.47 (s, 9 H, CO₂Bu^t), 2.90 (m, 1 H, HC-3), 4.79 (dd, 1 H, J = 7.9, 3.2, HC-2), 5.08 (s, 2 H, CH₂Ar), 5.46 (d, 1 H, J = 7.9, NH), 7.34 (s, 5 H, Ar), 9.76 (s, 1 H, CHO).

Reductive Amination Product (2S,3S,2'S)-18. To a stirred solution of (2S,3R)-17 (140 mg, 0.436 mmol) in 10 mL of dry MeOH was added a mixture of L-alanine methyl ester hydrochloride (300 mg, 2.18 mmol) and NaCNBH₃ (22 mg, 0.35 mmol) at room temperature. The homogeneous reaction mixture was stirred for 45 min, the solvent was evaporated, and the residue was partitioned between 25 mL of saturated NaHCO3 and 40 mL of Et₂O. The organic layer was dried and evaporated, and the residue was purified by filtration through a short column of silica gel (hexane/EtOAc, 2/1) to leave (2S,3S,2'S)-18 as a colorless oil in 89% yield: ¹H NMR δ 0.97 (d, 3 H, J = 6.9, CH₃C-3), 1.26 (d, $3 \text{ H}, J = 7.0, \text{CH}_3\text{C}-2'$), 1.46 (s, 9 H, CO_2Bu^t), 2.20 (m, 1 H, HC-3), 2.39 (dd, 1 H, J = 11.8, 7.5, HC-4), 2.62 (dd, 1 H, J = 11.8, 5.4, HC-4), 3.28 (q, 1 H, J = 7.0, HC-2'), 3.70 (s, 3 H, CO₂CH₃), 4.24 (dd, 1 H, J = 8.3, 4.2, HC-2), 5.11 (s, 2 H, CH₂Ar), 6.05 (d, 1 H, J)J = 8.3, NH), 7.36 (s, 5 H, Ar). Anal. Calcd for $C_{21}H_{32}N_2O_6$: C, 61.8; H, 7.9; N, 6.9. Found: C, 62.0; H, 7.7; N, 6.6.

(2S, 3R, 2'S)-18: ¹H NMR δ 0.83 (d, 3 H, J = 6.8, CH₃C-3), 1.30 (d, 3 H, J = 7.0, CH₃C-2'), 1.46 (s, 9 H, CO₂Bu^t), 2.20 (m, 1 H, HC-3), 2.30 (dd, 1 H, J = 12.2, 5.6, HC-4), 2.64 (dd, 1 H, J = 12.2, 8.7, HC-4), 3.34 (q, 1 H, J = 7.0, HC-2'), 3.70 (s, 3 H, CO₂CH₃), 4.45 (dd, 1 H, J = 8.8, 3.0, HC-2), 5.11 (s, 2 H, CH₂Ar), 5.72 (d, 1 H, J = 8.8, NH), 7.35 (s, 5 H, Ar).

 γ -Lactam Dipeptide (3S,4S,2'S)-19. A solution of (2S,3S,2'S)-18 (45 mg, 0.11 mmol) in formic acid (2.5 mL, 95–97%) was heated to 65–70 °C for 90 min. Excess formic acid was evaporated, and the residue was dried for 0.5 h at 50 °C (0.5 mmHg) (Kugelrohr). The solid residue was dissolved in 2 mL of DMF, and pyridine $(10 \ \mu\text{L}, 140 \ \text{mol} \ \%)$ was added. The clear solution was stirred at 55–60 °C for 90 min, the DMF was evaporated, the oily residue was dissolved in 30 mL Et₂O, and the organic layer was washed with 15 mL of saturated NaHCO₃

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and 15 mL of 1 M H₃PO₄, dried, and evaporated. The residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 2/3) to leave (3S,4S,2'S)-19 as a oil in 66% yield: ¹H NMR δ 0.98 (d, 3 H, J = 7.0, CH₃C-4), 1.42 (d, 3 H, J = 7.5, CH₃C-2'), 2.88 (m, 1 H, HC-4), 3.08 (d, 1 H, J = 9.6, H₆C-5), 3.54 (dd, 1 H, J = 9.5, 5.9, H_aC-5), 3.72 (s, 3 H, CO₂CH₃), 4.42 (t, 1 H, J = 5.8, HC-3), 4.84 (q, 1 H, J = 7.5, HC-2'), 5.13 (s, 2 H, CH₂Ar), 5.31 (d, br, 1 H, NH), 7.36 (s, 5 H, Ar). Anal. Calcd for C₁₇H₂₂N₂O₅: C, 61.1; H, 6.6; N, 8.4. Found: C, 61.2; H, 6.2; N, 8.4.

(3*S*,4*R*,2'*S*)-19: ¹H NMR δ 1.25 (d, 3 H, *J* = 6.4, CH₃C-4), 1.41 (d, 3 H, *J* = 7.5, CH₃C-2'), 2.27 (m, 1 H, HC-4), 2.95 (t, 1 H, *J* = 9.3, H_aC-5), 3.50 (dd, 1 H, *J* = 9.2, 8.1, H_bC-5), 3.71 (s, 3 H, CO₂CH₃), 4.08 (t, 1 H, *J* = 8.8, HC-3), 4.85 (q, 1 H, *J* = 7.5, HC-2'), 5.13 (s, 2 H, CH₂Ar), 5.19 (s, br, 1 H, NH), 7.35 (s, 5 H, Ar).

Cyclic Carbamate (4S,5R)-20. To a stirred solution of (2S,3R)-15 (66 mg, 0.35 mmol) and Et₃N (0.41 mL, 3.15 mmol) in 5 mL of CH₂Cl₂ was added COCl₂ (1.27 mL, 10 wt% COCl₂ in toluene) at 0 °C under N₂. The ice bath was removed, stirring was continued for 30 min, 10 mL of saturated NaHCO₃ and 10 mL of CH₂Cl₂ were added, and the layers were separated. The organic layer was dried and evaporated, and the residue was chromatographed (silica gel, hexane/EtOAc, 1/2) to leave (4S,5R)-20 as a white solid in 50% yield: mp 129-131 °C; ¹H NMR δ 1.20 (d, 3 H, J = 6.8, CH₃C-5), 1.50 (s, 9 H, CO₂Bu^t), 2.30 (m, 1 H, HC-5), 3.65 (dd, 1 H, J = 7.2, 1.3, HC-4), 3.94 (dd, 1 H, J = 11.1, 8.0, HC-6), 4.21 (dd, 1 H, J = 11.2, 3.7, HC-6), 5.7 (s, br, 1 H, NH). Anal. Calcd for C₁₀H₁₇NO₄: C, 55.8; H, 8.0; N, 6.5. Found: C, 56.8; H, 8.0; N, 6.3.

(48,58)-20: mp 118–120 °C; ¹H NMR δ 1.06 (d, 3 H, J = 7.0, CH₃C-5), 1.51 (s, 9 H, CO₂Bu¹), 2.48 (m, 1 H, HC-5), 4.11 (d, 1 H, J = 4.7, HC-5), 4.16 (dd, 1 H, J = 11.1, 3.5, HC-6), 4.32 (dd, 1 H, J = 11.1, 2.8, HC-6), 5.59 (s, 1 H, NH).

Dimethyl N-(9-Phenylfluorenyl)-3-methyl-L-aspartate (23). To a stirred solution of KHMDS (5.5 mL, 3.24 mmol, 180 mol %, 0.6 M in toluene) in 40 mL of dry THF was added 22 (720 mg, 1.8 mmol, prepared via dimethyl aspartate (21) as described ²¹) dissolved in 5 mL of THF drop-by-drop at under N_2 at -72 °C. The pale yellow solution was stirred at -75 °C for 1 h, and then CH₃I (0.57 mL, 9 mmol) in 1.3 mL of THF was added at -75 °C. The reaction mixture was stirred for 2 h at -75 °C, 2 mL of MeOH were added at -75 °C, the reaction mixture was partitioned between 40 mL of 1 M H_3PO_4 and 40 mL of Et_2O , and the water layer was extracted with Et_2O (2 × 40 mL). The combined organic layer was washed with 40 mL of saturated $Na_2S_2O_3$ and brine, dried, and evaporated, and the residue was purified by MPLC (silica gel, hexane/EtOAc, 8/1) to leave 23 (600 mg, 82%) as a mixture of diastereoisomers (3/2, according)to ¹H NMR): ¹H NMR δ 0.92 (d, J = 7.0, CH₃C-3), 1.46 (d, J = 7.0, CH₃C-3), 2.5-2.7 (m, 2 HC-3), 2.85-3.12 (m, br, 2 HC-2 and 2 NH), 3.17, 3.26, 3.48, 3.72 (4 s, CO₂CH₃) 7.17-7.43 (m, 11 H, Ar), 7.64-7.71 (m, 2 H, Ar). Anal. Calcd for C₂₆H₂₅NO₄: C, 75.2; H, 6.1; N, 3.4. Found: C, 75.1; H, 6.1; N, 3.4. As a side product, 117 mg (15%) of dimethylated product was isolated.

Methyl (2S)-2-[(9-Phenylfluorenyl)amino]-3-methyl-4hydroxybutanoate (24). To a stirred solution of 23 (580 mg, 1.4 mmol) in 50 mL of dry THF was added DIBAL (4.2 mL, 4.2 mmol, 1 M in hexane) dropwise at -35 °C under N₂. The reaction mixture was stirred at -30 °C for 1 h, it was poured into a stirred mixture of 50 mL of 1 M H₃PO₄ and 50 mL of Et₂O, and the phases were separated. The water layer was extracted with 30 mL of Et₂O, and the combined organic layer was dried and evaporated. The residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 2/1) to leave 508 mg (93%) of 24 as a mixture of diastereoisomers $(3/2, according to {}^{1}H NMR)$: ¹H NMR δ 0.53 (d, J = 7.0, CH₃C-3), 0.90 (d, J = 7.2, CH₃C-3), 1.65 (m, HC-3), 1.93 (m, HC-3), 2.48 (d, J = 9.9, HC-2), 2.76 (d, J = 3.8, HC-2), 3.19, 3.28 (2 s, CO₂CH₃), 3.29-3.63 (m, H₂C-4), 7.15-7.44 (m, 11 H, Ar), 7.67-7.71 (m, 2 H, Ar). Anal. Calcd for C25H25NO3: C, 77.5; H, 6.5; N, 3.6. Found: C, 77.4; H, 6.6; N, 3.6

Methyl (2S,3S)-2-[(9-Phenylfluorenyl)amino]-3-methyl-4-oxobutanoate [(2S,3S)-25] and Methyl (2S,3R)-2-[(9-Phenylfluorenyl)amino]-3-methyl-4-oxobutanoate [(2S,3R)-25]. To a stirred suspension of NCS (682 mg, 5.12 mmol) in 20 mL of toluene was added Me₂S (512 μ L, 6.35 mmol) at 0 °C under N₂. The suspension was cooled to -25 °C, and 24 (495 mg, 1.28 mmol) in 4 mL of toluene was added dropwise. Stirring was continued for 4 h at -25 °C, and then Et₃N (717 μ L, 5.12 mmol) in 2 mL of toluene was added. The cold bath was removed, and after 10 min, the reaction mixture was partitioned between 50 mL of 1 M H₃PO₄ and 50 mL of Et₂O. The organic layer was washed with 20 mL of H₂O, dried, and evaporated, and the diastereoisomeric aldehydes were purified by chromatography (low pressure, silica gel, hexane/EtOAc, 6/1) in 78% yield and separated by MPLC (silica gel, hexane/EtOAc, 16/1).

Major isomer (2S,3S)-25: ¹H NMR δ 1.06 (d, 3 H, J = 6.9, CH₃C-3), 2.35 (m, 1 H, HC-3), 3.0–3.2 (m, 2 H, HC-2, NH), 3.32 (s, 3 H, CO₂CH₃), 7.18–7.40 (m, 11 H, Ar), 7.66–7.73 (m, 2 H, Ar), 9.21 (s, 1 H, CHO). Anal. Calcd for C₂₅H₂₃NO₃: C, 77.8; H, 6.0; N, 3.6. Found: C, 77.8; H, 6.1; N, 3.6. Minor isomer (2S,3R)-25: ¹H NMR δ 0.81 (d, 3 H, J = 6.9,

Minor isomer (2S,3R)-25: ¹H NMR δ 0.81 (d, 3 H, J = 6.9, CH₃C-3), 2.42 (m, 1 H, HC-3), 2.90 (t, br, 1 H, J = 9.0, HC-2), 3.10 (d, br, 1 H, NH), 3.24 (s, 3 H, CO₂CH₃), 7.18–7.39 (m, 11 H, Ar), 7.67–7.74 (m, 2 H, Ar), 9.47 (d, 1 H, J = 3.2, CHO).

Reductive Amination Product (2S,3R,2'S)-26. To a stirred solution of (2S,3S)-25 (355 mg, 0.92 mmol) in 15 mL of dry MeOH was added a mixture of L-alanine methyl ester hydrochloride (640 mg, 4.6 mmol) and NaCNBH₃ (50 mg, 0.8 mmol) at room temperature. The pale yellow reaction mixture was stirred for 1 h, the solvent was evaporated, and the residue was partitioned between 40 mL of saturated NaHCO₃ and 50 mL of Et₂O. The water layer was extracted with 20 mL of Et₂O, and the combined organic layer was ashed with H₂O, dried, and evaporated. The residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 3/1) to give 417 mg (95%) of (2S,3R,2'S)-26 as a thick oil.

(2S,3R,2'S)-26: ¹H NMR δ 0.87 (d, 3 H, J = 6.8, CH₃C-3), 1.11 (d, 3 H, J = 7.0, CH₃C-2'), 1.58 (m, 1 H, HC-3), 2.19 (dd, 1 H, J = 12.2, 6.1, HC-4), 2.52 (dd, 1 H, J = 12.2, 7.3, HC-4), 2.60 (d, 1 H, J = 4.0, HC-2), 3.17 (q, 1 H, J = 7.0, HC-2'), 3.24 (s, 3 H, CO₂CH₃), 3.68 (s, 3 H, CO₂CH₃), 7.12–7.46 (m, 11 H, Ar), 7.64–7.70 (m, 2 H, Ar).

(2S,3S,2'S)-26: ¹H NMR δ 0.69 (d, 3 H, J = 6.9, CH₃C-3), 1.24 (d, 3 H, J = 6.9, CH₃C-2'), 1.72 (m, 1 H, HC-3), 1.99 (s, br, 1 H, NH), 2.41 (d, 1 H, J = 7.6, HC-2, and dd, 1 H, J = 11.3, 6.6, HC-4), 2.60 (dd, 1 H, J = 11.7, 5.6, HC-4), 3.19 (s, 3 H, CO₂CH₃), 3.28 (q, 1 H, J = 6.9, HC-2'), 3.74 (s, 3 H, CO₂CH₃), 7.14–7.43 (m, 11 H, Ar), 7.65–7.71 (m, 2 H, Ar).

 γ -Lactam Dipeptide (3S, 4R, 2'S)-27. A solution of (2S, 3R, 2'S)-26 (150 mg, 0.32 mmol) in 15 mL of toluene was refluxed at 120 °C bath temperature for 22 h. The solvent was evaporated, and the residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 3/1) to give 104 mg (75%; 95% based on recovered starting material) of (3S, 4R, 2'S)-27 as a thick oil.

(3S,4R,2'S)-27: ¹H NMR δ 0.17 (d, 3 H, J = 6.7, CH₃C-4), 1.27 (d, 3 H, J = 7.3, CH₃C-2'), 2.07 (m, 1 H, HC-4), 2.50 (d, 1 H, J = 8.6, HC-3), 2.52 (dd, 1 H, J = 9.3, 8.4, HC-5), 3.24 (s, br, NH), 3.32 (dd, 1 H, J = 9.3, 8.3, HC-5), 3.72 (s, 3 H, CO₂CH₃), 4.75 (q, 1 H, J = 7.4, HC-2'), 7.17–7.5 (m, 11 H, Ar), 7.64–7.73 (m, 2 H, Ar). Anal. Calcd for C₂₈H₂₈N₂O₃: C, 76.3; H, 6.4; N, 6.4. Found: C, 76.4; H, 6.5; N, 6.3.

(35,45,2'S)-27: ¹H NMR δ 1.09 (d, 3 H, J = 7.0, CH₃C-4), 1.23 (d, 3 H, J = 7.4, CH₃C-2'), 1.50 (m, 1 H, HC-4), 2.75 (d, 1 H, J = 6.7, HC-3), 2.77 (d, 1 H, J = 9.4, HC-5), 3.0 (dd, 1 H, J = 9.4, 5.4, HC-5), 3.69 (s, 3 H, CO₂CH₃), 4.73 (q, 1 H, J = 7.4, HC-2'), 7.1–7.5 (m, 1 H, Ar), 7.64–7.75 (m, 2 H, Ar).

(3S,4R,2'R)-27: ¹H NMR δ 0.13 (d, 3 H, J = 6.7, CH₃C-4), 1.37 (d, 3 H, J = 7.4, CH₃C-2'), 1.93 (m, 1 H, HC-4), 2.58 (d, 1 H, J = 7.8, HC-3), 2.62 (dd, 1 H, J = 8.4, 7.8, HC-5), 3.29 (t, 1 H, J = 8.8, HC-5), 3.61 (s, 3 H, CO₂CH₃), 4.83 (q, 1 H, J = 7.4, HC-2'), 7.17–7.49 (m, 11 H, Ar), 7.64–7.73 (m, 2 H, Ar).

(3S,4S,2'R)-27: ¹H NMR δ 0.99 (d, 3 H, J = 7.0, CH_oC-4), 1.30 (d, 3 H, J = 7.4, CH₃C-2'), 1.48 (m, 1 H, HC-4), 2.69 (d, 1 H, J = 9.7, HC-5), 2.79 (d, 1 H, J = 6.7, HC-3), 3.01 (s, 1 H, NH), 3.11 (dd, 1 H, J = 9.7, 5.2, HC-5), 3.58 (s, 3 H, CO₂CH₃), 4.80 (q, 1 H, J = 7.4, HC-2'), 7.15–7.47 (m, 11 H, Ar), 7.64–7.72 (m, 2 H, Ar).

Dimethyl 2-N-(9-Phenylfluorenyl)-3-ethyl-L-aspartate (28). To a stirred solution of KHMDS (8.65 mL, 5.2 mmol, 0.6

M in toluene) in 60 mL of dry THF was added 22 (1.6 g, 4.0 mmol) dissolved in 8 mL of THF dropwise at -75 °C under N₂. The pale yellow solution was stirred at -75 °C for 45 min, and then EtOTf (568 μ L, 4.4 mmol) was added neat at once at -75 °C. After 10 min the reaction was quenched with 3 mL of MeOH and partitioned between 40 mL of 1 M H₃PO₄ and 50 mL of Et₂O. The water layer was extracted with 40 mL of Et₂O, the combined organic layers were dried and evaporated, and the residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 8/1) to leave 6% of 22 and 82% of a mixture of 28 (78%; 3.5/1 mixture of diastereoisomers) and dialkylation product (4%). The diastereoisomers of 28 and the dialkylation product were separated by MPLC (silica gel, hexane/EtOAc, 15/1) to yield 84% of 28 based on recovered 22 (6%).

(2S,3S)-28: ¹H NMR δ 0.77 (t, 3 H, J = 7.4, H₃C-2′), 1.56–1.79 (m, 2 H, H₂C-1′), 2.37 (m, 1 H, HC-3), 2.79 (dd, 1 H, J = 10.2, 6.6, HC-2), 3.04 (d, 1 H, J = 10.2, NH), 3.24 (s, 3 H, CO₂CH₃), 3.51 (s, 3 H, CO₂CH₃), 7.15–7.43 (m, 11 H, Ar), 7.63–7.70 (m, 2 H, Ar). Anal. Calcd for C₂₇H₂₇NO₄: C, 75.5; H, 6.3; N, 3.3. Found: C, 75.5; H, 6.2; N, 3.1.

(2S,3R)-28: ¹H NMR δ 0.74 (t, 3 H, J = 7.4, H₃C-2'), 1.14 (m, 1 H, HC-1'), 1.41 (m, 1 H, HC-1'), 2.47 (m, 1 H, HC-3), 2.85 (dd, 1 H, J = 11.4, 8.8, HC-2), 3.00 (d, 1 H, J = 11.5, NH), 3.13 (s, 3 H, CO₂CH₃), 3.76 (s, 3 H, CO₂CH₃e, 7.12–7.38 (m, 11 H, Ar), 7.65–7.71 (m, 2 H, Ar).

Methyl (2S,3S)-2-[(9-Phenylfluorenyl)amino]-3-ethyl-4hydroxybutanoate [(2S,3S)-29]. To a solution of (2S,3S)-28 (200 mg, 0.47 mmol) in 25 mL of toluene was added DIBAL (1.4 mL, 1.4 mmol, 1 M solution in hexane) at -49 °C drop-by-drop. The reaction mixture was stirred at -47 °C for 20 min, and then quenched by adding a mixture of 25 mL of 1 M H₃PO₄ and 30 mL of Et_2O at -47 °C. The phases were separated, and the organic layer was dried and evaporated to a residue of ca. 5 mL. This residue was filtered through a short column of silica gel (EtOAc), and the solvent was evaporated. Crude (2S,3S)-29 was dried for 45 min at 40 °C (0.5 mmHg) (Kugelrohr) and used immediately for the next oxidation step: mass recovery 84%; ¹H NMR δ 0.80 (t, 3 H, J = 7.0, H_3C-2'), 1.22–1.52 (m, 2 H, H_2C-1'), 2.81 (d, 1 H, J = 3.4, HC-2), 3.28 (s, 3 H, CO₂CH₃), 3.42 (dd, 1 H, J = 11.3, 2.0, HC-4), 3.60 (dd, 1 H, J = 11.3, 5.5, HC-4), 7.14–7.46 (m, 11 H, Ar), 7.67–7.70 (m, 2 H, Ar).

Methyl (2S,3S)-2-[(9-Phenylfluorenyl)amino]-3-ethyl-4oxobutanoate [(2S,3S)-30]. To a stirred suspension of NCS (200 mg, 1.5 mmol) in 10 mL of dry toluene was added Me_2S (150 μ L, 2.04 mmol) at 0 °C under N₂. The suspension was cooled to -25 °C, and crude (2S,3S)-29 (157 mg, 0.39 mmol) in 2 mL of toluene was added drop-by-drop. Stirring was continued for 4 h at -25 °C, and then Et₃N (208 μ L, 1.5 mmol) in 0.5 mL of toluene was added. The cold bath was removed, and after 10 min the reaction mixture was partitioned between 25 mL of 1 M H₃PO₄ and 30 mL of Et₂O. The organic layer was dried and evaporated. Purification of the residue by chromatography (low pressure, silica gel, hexane/EtOAc, 6/1) afforded (2S,3S)-30 in 59% yield based on (2S,3S)-28: ¹H NMR δ 0.78 (t, 3 H, J = 7.5, H₃C-2'), 1.50–1.60 (m, 1 H, HC-1'), 1.70-1.81 (m, 1 H, HC-1'), 2.20 (m, 1 H, HC-3), 2.98 (dd, 1 H, J = 9.0, 4.7, HC-2), 3.07 (d, 1 H, J = 10, NH), 3.29 (s, 3 H, CO₂CH₃), 7.17-7.41 (m, 11 H, Ar), 7.66-7.72 (m, 2 H, Ar), 9.35 (d, 1 H, J = 2.0, CHO).

Reductive Amination Product (2S,3R,2'S)-31. To a stirred solution of (2S,3S)-30 (100 mg, 0.25 mmol) in 7 mL of dry MeOH was added a mixture of L-alanine methylester hydrochloride (175 mg, 1.25 mmol) and NaCNBH₃ (14 mg, 0.22 mmol) at room temperature. The reaction mixture was stirred at room temperature for 3 h, the solvent was evaporated, and the residue was partitioned between 30 mL of saturated NaHCO3 and 40 mL of Et_2O . The H₂O layer was extracted with 30 mL of Et_2O , and the combined organic layer was dried and evaporated. Purification of the residue by chromatography (low pressure, silica gel, hexane/EtOAc, 4/1) left (2S, 3R, 2'S)-31 in 64% yield as a thick oil: ¹H NMR δ 0.67 (t, 3 H, J = 7.2, H₃C-2"), 1.13 (d, 3 H, J = 7.0, $CH_{3}C-2'$), 1.20–1.50 (m, 3 H, HC-3, $H_{2}C-1''$), 2.26 (dd, 1 H, J =12.3, 4.3, HC-4), 2.58 (dd, 1 H, J = 12.3, 7.0, HC-4), 2.66 (d, 1 H, J = 3.6, HC-2), 3.18 (q, 1 H, J = 7.0, HC-2'), 3.24 (s, 3 H, CO₂CH₃), 3.69 (s, 3 H, CO₂CH₃), 7.15–7.45 (m, 11 H, Ar), 7.64–7.70 (m, 2 H, Ar).

γ-Lactam Dipeptide (3S, 4R, 2'S)-32. A solution of 70 mg of (2S, 3R, 2'S)-31 in 10 mL of *p*-xylene was refluxed at 140–150 °C for 3 h. After evaporation of the solvent, (3S, 4R, 2'S)-32 was isolated in 86% yield by MPLC (silica gel, hexane/EtOAc, 5/1) as a white foam: ¹H NMR δ 0.38 (m, 4 H, H₃C-2''), 0.59 (m, 1 H, HC-1''), 1.29 (d, 3 H, J = 7.4, CH₃C-2'), 1.93 (m, 1 H, HC-4), 2.59 (d, 1 H, J = 8.5, HC-3), 2.61 (dd, 1 H, J = 9.5, 7.6, HC-5), 3.16 (s, 1 H, NH), 3.37 (t, 1 H, J = 8.8, HC-5), 3.73 (s, 3 H, CO₂CH₃), 4.76 (q, 1 H, J = 7.5, HC-2'), 7.16–7.54 (m, 11 H, Ar), 7.63–7.71 (m, 2 H, Ar). Anal. Calcd for C₂₉H₃₀N₂O₃: C, 76.6; H, 6.7; N, 6.2. Found: C, 76.6; H, 6.7; N, 6.1.

2-N-(9-Phenylfluorenyl)-3-isopropyl-L-aspartic Acid α tert-Butyl &-Methyl Diester [(2S,3R)- and (2S,3S)-33]. To a stirred solution of KHMDS (0.43 mL, 0.26 mmol, 0.6 M in toluene) in 5 mL of dry THF was added 5 (50 mg, 0.13 mmol) in 1 mL of THF drop-by-drop at -78 °C. After 45 min, i-PrOTf (0.4 mL, 0.26 mmol, 0.67 M in hexane) was added at -78 °C, and after 45 min another 0.43-mL portion of KHDMS was added. followed by 0.4 mL of i-PrOTf in hexane after 15 min. The reaction was stirred at -78 °C for 60 min and then quenched with 15 mL of 1 M H_3PO_4 and 20 mL of Et_2O . The phases were separated, the water laver was extracted with 20 mL of Et₂O, the combined organic layer was dried and evaporated, and the residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 12/1), affording 27.2 mg (43%) of 33 as a thick oil (ratio of diastereoisomers 5/1 by ¹H NMR) and 25.1 mg (45%) of recovered 5. The diastereoisomers of 33 are separable by MPLC (silica gel, hexane/CHCl₃/EtOAc, 10/20/0.6).

Major isomer (2S,3S)-**33**: ¹H NMR δ 0.41 (d, 3 H, J = 6.5, CH₃C-1'), 0.67 (d, 3 H, J = 6.5, CH₃C-1'), 1.24 (s, 9 H, CO₂Bu^t), 1.96 (m, 1 H, HC-1'), 2.14 (dd, 1 H, J = 10, 4.9, HC-3), 2.66 (d, 1 H, J = 4.9, HC-2), 3.35 (br, 1 H, NH), 3.68 (s, 3 H, CO₂CH₃), 7.17–7.42 (m, 11 H, Ar), 7.65–7.70 (m, 2 H, Ar). Anal. Calcd for C₃₁H₃₅NO₄: C, 76.7; H, 7.3; N, 2.9. Found: C, 76.6; H, 7.3; N, 2.8.

Minor isomer (2S,3R)-33: ¹H NMR δ 0.54 (d, 3 H, J = 6.6, CH₃C-1'), 0.83 (d, 3 H, J = 6.6, CH₃C-1'), 1.10 (s, 9 H, CO₂Bu^t), 2.05 (m, 1 H, HC-1'), 2.24 (dd, 1 H, J = 8.0, 6.0, HC-3), 2.9 (d, 1 H, J = 5.3, HC-2), 3.10 (br, 1 H, NH), 3.71 (s, 3 H, CO₂CH₃), 7.13-7.41 (m, 11 H, Ar), 7.63-7.68 (m, 2 H, Ar).

tert-Butyl (2S,3S)-2-[(9-Phenylfluorenyl)amino]-3-isopropyl-4-hydroxybutanoate [(2S,3S)-34]. To a stirred solution of (2S,3S)-33 (220 mg, 0.45 mmol) in 15 mL of toluene was added DIBAL (1.4 mL, 1.4 mmol, 1 M in hexane) at -30 °C. The reaction mixture was stirred at -30 °C for 20 min and then quenched by adding a mixture of 20 mL of 1 M H₃PO₄ and 30 mL of EtO₂ at -30 °C. The phases were separated, the water layer was extracted with 25 mL of Et₂O, and the combined organic layer was dried and evaporated to a residue of ca. 3 mL, which was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 7/1) to give 161 mg (78%) of (2S,3S)-34 as a colorless oil: ¹H NMR δ $0.52 (d, 3 H, J = 6.7, CH_3C-1'), 0.82 (d, 3 H, J = 6.7, CH_3C-1'),$ 1.21 (s, 9 H, CO₂Bu^t), 1.2-1.35 (m, 1 H, HC-1'), 1.5-1.62 (m, 1 H, HC-3), 2.77 (d, 1 H, J = 3.8, HC-2), 3.61 (dd, 1 H, J = 11.4, 8.0, HC-4), 3.77 (dd, 1 H, J = 11.4, 3.1, HC-4), 7.18-7.42 (m, 11 H, Ar), 7.67-7.71 (m, 2 H, Ar). Anal. Calcd for C₃₀H₃₅NO₃: C, 78.7; H, 7.7; N, 3.1. Found: C, 78.7; H, 7.8; N, 3.0.

tert-Butyl (2S,3S)-2-[(Benzoxycarbonyl)amino]-3-isopropyl-4-hydroxybutanoate [(2S,3S)-35]. A solution of (2S,3S)-34 (175 mg, 0.38 mmol) in 12 mL of MeOH/HOAc (15/1) was hydrogenolyzed overnight under 1 atm of H₂ using 80 mg Pd/C (10%) as the catalyst. The reaction mixture was filtered through Celite, the catalyst was washed with MeOH, the filtrate and washings were evaporated, and the residue was partitioned between 15 mL of H₂O and 20 mL of Et₂O. The organic layer was extracted with 10 mL of 0.5 M H₃PO₄, and the combined water layer was adjusted to pH 9 with saturated Na₂CO₃ and extracted with CHCl₃ (3 × 20 mL). The organic layer was dried and evaporated to give crude amine in ca. 85% yield.

To a stirred solution of 57 mg (0.26 mmol) of crude amine and 140 mg (1.3 mmol) of NaHCO₃ in 8 mL of H₂O/EtOAc, 1/1, was added CBZ-Cl (60 μ L, 0.39 mmol) in 1 mL of EtOAc dropwise at 0 °C. The mixture was stirred at 0 °C for 90 min, the phases were separated, and the water layer was extracted with 20 mL of EtOAc. The combined organic layer was dried and evaporated, and the residue was purified by chromatography (low pressure,

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silica gel, hexane/EtOAc, 9/2) to give 60 mg (66%) of (2S,3S)-35 as a colorless oil and 22 mg (30%) of lactone (3S,4S)-36.

(2S,3S)-35: ¹H NMR δ 0.88 (d, 3 H, J = 6.8, CH₃C-1'), 0.97 (d, 3 H, J = 6.8, CH₃C-1'), 1.47 (s, 9 H, CO₂Bu^t), 1.68 (m, 1 H, HC-1'), 2.0 (m, 1 H, HC-3), 3.47 (dd, br, 2 H, J = 11.0, H₂C-4), 3.73 (s, br, 1 H, OH), 4.62 (dd, 1 H, J = 8.0, 3.1, HC-2), 5.12 (s, 2 H, CH₂Ar), 5.68 (d, 1 H, J = 7.7, NH), 7.31–7.37 (s, 5 H, Ar). Anal. Calcd for C₁₉H₂₉NO₅: C, 64.9; H, 8.3; N, 4.0. Found: 64.9; H, 8.1; N, 4.0.

(3S,4S)-36: mp 81-83 °C; ¹H NMR δ 0.94 (d, 3 H, J = 6.5, CH₃C-1'), 1.00 (d, 3 H, J = 6.6, CH₃C-1'), 1.85 (m, 1 H, HC-1'), 2.33 (m, 1 H, HC-4), 3.93 (t, 1 H, J = 10, HC-5), 4.26 (t, 1 H, J = 10, HC-5), 4.40 (t, 1 H, J = 8.5, HC-3), 5.14 (m, 2 H, CH₂Ar), 5.26 (d, 1 H, J = 8.5, NH), 7.31-7.36 (s, 5 H, Ar). Anal. Calcd for C₁₅H₁₉NO₄: C, 65.0; H, 6.9; N, 5.0. Found: C, 64.9; H, 7.0; N, 4.9.

tert-Butyl (2S,3S)-2-[(Benzoxycarbonyl)amino]-3-isopropyl-4-oxobutanoate [(2S,3S)-37]. To a stirred suspension of NCS (77 mg, 0.57 mmol) in 4.5 mL of dry toluene was added Me_2S (58 µL, 0.72 mmol) at 0 °C under N₂. The suspension was cooled to -25 °C and (2S,3S)-35 (45 mg, 0.128 mmol) in 0.5 mL of toluene was added dropwise. Stirring was continued for 4 h at -25 °C, and then Et₃N (80 μ L, 0.57 mmol) in 0.5 mL of toluene was added. The cold bath was removed, and after 10 min, the reaction was partitioned between 20 mL of 1 M $\rm H_3PO_4$ and 25 mL of Et_2O . The organic layer was washed three times with 20 mL of H₂O, dried, and evaporated. The residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 5/1) to give (2S,3S)-37 in 80% yield, based on 14% recovered (2S,3S)-35: ¹H NMR δ 1.05 (d, 3 H, J = 6.9, CH₃C-1'), 1.09 (d, 3 H, J = 6.9, CH₃C-1'), 1.45 (s, 9 H, CO₂Bu^t), 2.16 (m, 1 H, HC-1'), 2.45 (m, 1 H, HC-3), 4.62 (dd, 1 H, J = 8.3, 6.0, HC-2), 5.11 (s, 2 H, CH₂Ar), 5.45 (d, 1 H, J = 8.3, NH), 7.31–7.36 (s, 5 H, Ar), 9.72 (d, 1 H, J = 2.9, CHO). Anal. Calcd for $C_{19}H_{27}NO_5$: C, 65.3; H, 7.8; N, 4.0. Found: C, 64.8; H, 7.7; N, 4.0.

Reductive Amination Product (2S,3R,2'S)-38. To a stirred solution of (2S,3S)-37 (20 mg, 0.057 mmol) in 2 mL of dry MeOH was added a mixture of L-alanine methyl ester hydrochloride (50 mg, 0.36 mmol) and NaCNBH₃ (6 mg, 0.1 mmol) at room temperature. After 2 h, the solvent was evaporated, and the residue was partitioned between 10 mL of saturated NaHCO₃ and 20 mL of Et₂O. The water layer was extracted with 10 mL of Et₂O, the combined organic layer was dried and evaporated, and the residue was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 4/1) to give 13 mg (52%) of (2S,3R,2'S)-38 as a thick oil: ¹H NMR δ 0.91 (d, 3 H, J = 6.3, CH₃C-1"), 1.03 (d, 3 H, J= 6.1, CH_3C-1''), 1.27 (d, 3 H, J = 7.0, CH_3C-2'), 1.46 (s, 9 H, CO₂Bu^t), 1.62-1.79 (m, 2 H, HC-1", HC-3), 2.54-2.70 (m, 2 H, $H_2\bar{C}-4$), 3.34 (q, 1 H, J = 7.0, HC-2'), 3.70 (s, 3 H, CO₂CH₃), 4.56 $(dd, 1 H, J = 8.7, 2.5, HC-2), 5.11 (s, 2 H, CH_2Ar), 6.48 (d, 1 H, J)$ J = 8.6, NH), 7.27–7.37 (s, 5 H, Ar).

 γ -Lactam Dipeptide (3S,4R,2'S)-39. A solution of (2S,3R,2'S)-38 (12 mg, 0.027 mmol) in formic acid (1.5 mL, 95-97%) was heated to 70 °C for 90 min. Excess formic acid was evaporated, and the residue was dried for 15 min at 40 °C (0.5

mmHg) (Kugelrohr). The oily residue was dissolved in 1.5 mL of DMF and two drops of pyridine, the clear solution was stirred at 65 °C for 2.5 h, and the solvent was evaporated. The residue was dissolved in 15 mL of Et₂O, washed with 10 mL of saturated NaHCO₃ and 10 mL of 1 M H₃PO₄, dried, and evaporated, leaving a residue that was purified by chromatography (low pressure, silica gel, hexane/EtOAc, 3/2) to give (3S,4R,2'S)-**39** as an oil in 62% yield: ¹H NMR δ 0.97 (d, 3 H, J = 7.0, CH₃C-1″), 0.99 (d, 3 H, J = 7.0, CH₃C-1″), 0.99 (d, 3 H, J = 7.0, CH₃C-1″), 1.42 (d, 3 H, J = 7.5, CH₃C-2′), 1.86–1.94 (m, 1 H, HC-1″), 2.07–2.15 (m, 1 H, HC-4), 3.04 (t, 1 H, J = 9.2, H_αC-5), 3.44 (t, 1 H, J = 9.0, H_βC-5), 3.72 (s, 3 H, CO₂CH₃), 4.29 (dd, 1 H, J = 9.4, NH), 5.14 (s, 2 H, CH₂Ar), 7.29–7.36 (s, 5 H, Ar). Anal. Calcd for C₁₉H₂₆N₂O₅: C, 63.0; H, 7.2; N, 7.7. Found: C, 62.7; H, 7.3; N, 7.5.

Preparation of Isopropyl Triflate in Hexane. A solution of 0.69 mL of i-PrOH (distilled from CaH₂) and 0.74 mL of pyridine (distilled from CaH₂) in 4.5 mL of hexane (distilled from NaH) was added over a period of 20 min to a stirred solution of 1.51 mL of $(Tf)_2O$ in 7.5 mL of hexane at 0 °C. The reaction mixture was stirred at 0 °C for 15 min and then filtered through a short pad of Na₂SO₄ to give a colorless, clear solution, 0.67 M in hexane, which was used immediately.

Acknowledgment. We thank Dr. P. Gmeiner for helpful discussions and E. Civitello for the ¹H NMR NOESY experiments. J.-P.W. is a fellow of the Schweizerischer Nationalfonds zur Förderung der Wissenschaftlichen Forschung.

Registry No. 5, 120230-41-3; (2S,3S)-6, 120230-42-4; (2S,3R)-6, 120230-43-5; (2S,3S)-7, 120230-46-8; (2S,3R)-7, 120230-47-9; (2S,3R)-8, 120230-48-0; (2S,3S,2'S)-9, 120230-49-1; (2S,3S,2'S)-11, 120230-50-4; (2S,3S,2'S)-12, 120230-51-5; (3S,4S,2'S)-13, 120230-52-6; 14, 120230-53-7; (2S,3R)-15, 120230-54-8; (2S,3S)-15, 120230-55-9; (2S,3R)-16, 120262-55-7; (2S,3S)-16, 120230-56-0; (2S,3R)-17, 120230-57-1; (2S,3S)-17, 120230-58-2; (2S,3S,2'S)-18, 120230-59-3; (2S,3R,2'S)-18, 120329-59-1; (3S,4S,2'S)-19, 120230-60-6; (3S,4R,2'S)-19, 120328-49-6; (4S,5R)-20, 120230-61-7; (4S,5S)-20, 120262-56-8; 22, 120230-62-8; (S,3S)-23, 120230-63-9; (2S,3R)-23, 120230-64-0; (2S,3S)-24, 120230-68-4; (2S,3R)-24, 120230-69-5; (2S,3S)-25, 120230-71-9; (2S,3R)-25, 120230-72-0; (2S, 3R, 2'S)-26, 120230-74-2; (2S, 3S, 2'S)-26, 120328-50-9; (2S, 3R, 2'R)-26, 120328-51-0; (2S, 3S, 2'R)-26, 120328-52-1; (3S,4R,2'S)-27, 120230-76-4; (3S,4S,2'S)-27, 120328-53-2; (3S,4R,2'R)-27, 120328-54-3; (3S,4S,2'R)-27, 120328-55-4; (2S,3S)-28, 120230-65-1; (2S,3R)-28, 120230-66-2; (2S,3S)-29, 120230-70-8; (2S,3S)-30, 120230-73-1; (2S,3R,2'S)-31, 120230-75-3; (3S,4R,2'S)-32, 120230-77-5; (2S,3S)-33, 120230-44-6; (2S,3R)-33, 120230-45-7; (2S,3S)-34, 120230-78-6; (2S,3S)-35, 120230-79-7; (3S,4S)-36, 120230-80-0; (2S,3S)-37, 120230-81-1; (2S,3R,2'S)-38, 120230-82-2; (3S,4R,2'S)-39, 120230-83-3; H-Ala-OMe+HCl, 2491-20-5; L-(PhFl)NHCH(CO₂Me)CMe₂CO₂Me, 120230-67-3; H-D-Ala-OMe-HCl, 14316-06-4.